General PCR set up for genomic DNA, Plasmid DNA and/or RT-PCR

Make up PCR mix for your desired product and number of reactions

a. PCR mix per reaction
   i. 2.5ul of 10X Hotstart Taq Buffer (Qiagen)
   ii. 0.5ul 10mM dNTP mix
   iii. Forward Primer of 0.15ul 20uM
   iv. Reverse Primer of 0.15ul 20uM
   v. 1ul 25mM MgCl2
   vi. H2O up to 25 ul total
   vii. Taq Polymerase (Qiagen) 0.125ul per reaction

- add 24 ul mix to each PCR reaction tube

-**FOR GENOMIC DNA** = add 1 ul of DNA sample to each tube (this amount may vary depending on the type of DNA you are using for PCR

-**FOR PLASMID DNA** = add 10-20 ng per of DNA per PCR reaction

-**FOR RT-PCR** start with 2 ul of RT reaction per PCR reaction

Set PCR machine for the desired settings according to your primers and product size.

Step 1	95 degrees	15 min
Step 2	95 degrees	1 min
Step 3	X degrees	1 min (annealing temp should be about 5 degrees lower than primer Tm)
Step 4	72 degrees	1 min per kb product size
	Decide the cycle number desired and repeat step 2-4 that number of times	
Step 5	72 degrees	10 min
Step 6	4 degrees until you can take it out.	

Run on an agarose gel and look for the appropriately sized bands.